



Journal of Applied Animal Research

ISSN: (Print) (Online) Journal homepage: https://www.tandfonline.com/loi/taar20

# Characterization of casein variants in the Guaymi and Guabala breeds through a low-density chip arrangement

Axel Villalobos-Cortés, Ginnette Rodriguez, Hilda Castillo & Selma Franco

To cite this article: Axel Villalobos-Cortés, Ginnette Rodriguez, Hilda Castillo & Selma Franco (2023) Characterization of casein variants in the Guaymi and Guabala breeds through a low-density chip arrangement, Journal of Applied Animal Research, 51:1, 69-73, DOI: 10.1080/09712119.2022.2154216

To link to this article: <u>https://doi.org/10.1080/09712119.2022.2154216</u>

0

© 2023 The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group



Published online: 11 Jan 2023.

Submit your article to this journal 🗹



View related articles



View Crossmark data 🗹

Taylor & Francis

OPEN ACCESS Check for updates

# Characterization of casein variants in the Guaymi and Guabala breeds through a low-density chip arrangement

Axel Villalobos-Cortés <sup>a</sup>, Ginnette Rodriguez<sup>b</sup>, Hilda Castillo<sup>c</sup> and Selma Franco<sup>d</sup>

<sup>a</sup>Laboratory of Applied Molecular Biology and Analysis, IDIAP, Panama City, Panama; <sup>b</sup>El Ejido Experimental Station, IDIAP, Panama City, Panama; <sup>c</sup>Technological University, Panama City, Panama; <sup>d</sup>Animal Health Laboratory, IDIAP, Divisa, Panama City, Panama

#### ABSTRACT

Studies of the genetic diversity of the Guaymi and Guabala cattle breeds have shown the need to evaluate various components, including the polymorphism of casein genes. The objective of this work is to characterize the casein variants in the Guaymi and Guabala landraces by means of a low-density SNP arrangement. Twenty-four SNP markers were typed in samples of Guabala and Guaymi Creole cattle. The values of  $H_{or}$   $H_{er}$  and  $F_{is}$ , considering only the polymorphic loci in the Guabala breed, were 0.438, 0.449, and 0.011, respectively. In the case of the Guaymi breed,  $H_{or}$ ,  $H_{er}$ , and  $F_{is}$  at the polymorphic loci were 0.513, 0.405, and -0.281, respectively. The effective number of alleles obtained from the Guabala breed was 1.167, and that in Guaymi was 1.257. This study determined the genetic diversity of the casein group in the Guabala race. Both breeds had high frequencies of the A2A2 genotype at rs43703011 (*CSN2*), which is considered favourable for production of quality milk. The identified markers will allow the design of strategies to reduce the levels of inbreeding and better understand the aptitudes of both breeds in terms of productivity.

## ARTICLE HISTORY

Received 12 July 2022 Accepted 28 November 2022

#### **KEYWORDS**

Livestock; local breeds; molecular genetics; biodiversity; biotechnology

# Introduction

Bovine milk is composed of 3-5% protein and consists of two main groups, whey proteins and caseins (Alim et al. 2014). Caseins represent 80% of dairy proteins in cattle (Martien et al. 1994), while serum proteins represent 14% (Roginski et al., 2003). The casein group is the largest among the components of milk and is encoded by four genes (Asmarasari et al. 2020): CSN1S1 (as1-casein) contributes 39-46% of the total of this group, CSN1S2 (as2-casein) contributes 8-11%, CSN2 (β-casein) contributes 25–35%, and CSN3 (kappa-casein) contributes 8-15% (Eigel et al. 1984; Ferretti et al. 1990; Roginski 2003). These genes are near each other on chromosome 6, within a range of 251.26 kb, which close relationship has suggested that the genes are often not inherited independently but as a single haplotype (Meier et al. 2019) in cattle as well as goats and sheep (Ferretti et al. 1990; Hayes et al. 1993; Popescu et al. 1996).

The genes of casein are closely related to the quality and productivity of milk and its derivatives. *CS1NS1* is associated with high milk production, as well as protein content (Eenennaam and Medrano 1991). *CSN2* has particular importance since it is related not only to the high yield and quality of milk (Kučerova et al. 2006) but also to a healthier product, particularly its A2A2 variant, unlike the A1A1 variant, which has been associated with the bioactive peptide  $\beta$ -casomorphin and human health risk factors such as ischaemic heart disease, arteriosclerosis, type I diabetes, sudden infant death

syndrome, and autism (Kaminski et al. 2007; Kost et al. 2009; Cieslinska et al. 2012). CSN1S2 is associated with protein yield (Nilsen et al. 2009). CSN3 is related to milk protein coding, the latter very important in the stability of the structure of casein micelles, milk production, and cheese quality (Alexander et al. 1988; Alim et al. 2014). Laible et al. (2016) revealed that milk protein genes have the potential to be used to improve the bovine milk component. The studies of genetic diversity that have been carried out in the Guaymi and Guabala breeds (Delgado et al. 2011; Ginja et al. 2019; Villalobos-Cortes et al. 2021a) have highlighted the need to evaluate various factors of productivity and milk quality, such as the variability of casein genes. The objective of this study is to characterize the caseins in the genomes of the Guaymi and Guabala breeds of Panama by genotyping single-nucleotide polymorphisms (SNPs).

#### **Materials and methods**

#### Sample collection

The polymorphism of 24 SNP markers of the caseins *CSN1S1* (2), *CSN2* (8), *CSN1S2* (2), and *CSN3* (12) of 34 samples of Criollo Guabala (15) and Guaymi cattle (19). The animals were selected within the conservation centres through a previously carried out genetic characterization, to guarantee the purity of both breeds (Villalobos-Cortes et al. 2020). These SNPs were selected from an array of 10,000 SNP markers in an Affymetrix Axiom

CONTACT Axel Villalobos-Cortés 🖾 villalobos.axel@gmail.com 🗈 Laboratory of Applied Molecular Biology and Analysis, IDIAP, City of Knowledge, Via Jacinto Palacios, Clayton Bldg 221, Panama

 $\ensuremath{\textcircled{O}}$  2023 The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

OrcunSNP Array platform, as part of the Innovative Management of Animal Genetic Resources (IMAGE) project in the Horizon 2020 framework programme.

#### **Genomic DNA isolation**

Five-millilitre samples of venous blood were taken from each animal and kept cold until arrival at the laboratory. DNA was extracted using the commercial kit DNeasy Blood and Tissue from Qiagen, obtaining an average concentration of 45 ng/ml and a volume of 50 µl per sample, with a total amount of 2.5 µg of DNA. Affymetrix analyses complied with the Nagoya protocol. Of the 10,000 SNPs selected, 8416 met the criteria recommended by the company, with a conversion threshold of 0.6. All SNPs were aligned with the UMD 3.1.1 reference genome (Elsik et al. 2016). The results obtained in VCF format were validated and transformed into GDA format using the program PGDSpider 2.1.1.5 (Lischer and Excoffier 2012), then converted to text and Excel formats. To verify the positions of the SNPs, the Integrative Genome Viewer program IGV 2.9.4.03 (Robinson et al. 2011) was used along with the Genome Data Viewer of the National Center for Biotechnology Information (NCBI), with the same reference genome, UMD 3.1.1. SNPs that had a reference number (RefSNP) were used to locate them in the reference genome position ARS.UCD.1.2 by Ensembl! (Howe et al. 2021) and the European Variation Archive (Cezard et al. 2021).

#### Genetic diversity analysis

To evaluate the genetic variability within each population, the following parameters were calculated: percentage of polymorphic loci, observed heterozygosity  $(H_{o})$ , expected heterozygosity  $(H_e)$ , effective number of alleles  $(N_e)$ , and deviations from Hardy-Weinberg (HW) equilibrium in each population, calculated by the exact test using the Markov chain method with a chain length of 1,000,000 and 100,000 memorization steps (Guo and Thompson 1992). Gene and genotypic frequencies and  $F_{is}$ ,  $F_{st}$ , and  $F_{it}$  values were also calculated (Wright 1965; Weir and Cockerham 1984). GENETIX 4.02 (Belkhir et al. 2003), GenAlEx 6.501 (Peakall and Smouse 2012) and ARLEQUIN 3.5. (Excoffier et al. 2005), the Shannon diversity index was calculated using GenAlEx 6.501. The polymorphic variants were subjected to Cattle QTLdb (Zhi-Liang et al. 2007) to identify possible associations with economic utility traits.

**Table 1.** Mean Shannon index (*I*) observed heterozygosity ( $H_o$ ), expected heterozygosity ( $H_e$ ) and HW equilibrium of casein gene variants of the Guaymi and Guabala breeds.

Gene	Variant (RefSNP)	Ι	H <sub>o</sub>	H <sub>e</sub>	HW				
CSN1S1	rs133474041	0.562	0.516	0.376	*				
CSN2	rs109299401	0.257	0.211	0.166	ns				
CSN2	rs43703011	0.218	0.158	0.133	ns				
CSN1S2	rs441966828	0.218	0.158	0.133	ns				
CSN3	rs450402006	0.611	0.370	0.421	ns				
CSN3	rs43703015	0.689	0.575	0.496	ns				
CSN3	rs43703016	0.688	0.556	0.495	ns				
CSN3	rs439304887	0.218	0.158	0.133	ns				
CSN3	rs110014544	0.688	0.594	0.495	ns				

#### Results

Of the 24 markers analysed, 23 were considered usable; in the Guabala breed, 20.83% of polymorphic loci were obtained (5), and in the Guaymi breed, 37.50% of polymorphic loci were obtained (8). Most of the variants, except for one belonging to the CSN2 gene (6:87183034) and three belonging to the CSN3 gene (6:87390198, 6:87390448 and 6:87390604), were identified. The location in the reference genome (UMD 3.1.1, Genome Data Viewer of NCBI). The values of  $H_o$ ,  $H_e$  and  $F_{is}$  (Table 1) considering only the polymorphic loci in the Guabala breed were 0.438, 0.449, and 0.0108, respectively. In the case of the Guaymi race,  $H_o$ ,  $H_e$  and  $F_{is}$ , also considering the polymorphic loci, were 0.513, 0.405, and -0.281, respectively. The  $N_e$  obtained from the Guabala breed was 1.167, and the Guaymi was 1.257, both considered low.

The general mean of the Shannon index considering the polymorphic loci was 0.173 for each population was 0.130 in Guabala and 0.215 in Guaymi. In the Guabala breed, most of the markers showed heterozygote deficits but did not reflect deviations from HW equilibrium. An excess of heterozygotes was obtained at all the markers evaluated in the Guaymi breed, with a greater difference being observed in rs133474041 (p < .05).

The variants of the genes that showed a high percentage of monomorphic alleles in both Guabala (79%) and Guaymi (63%) were, for the CSN1S1 gene, rs433385179 (G); for the CSN2 gene, rs433954503 (G), rs454083280 (T), rs43703013 (G), rs43703012 (G), rs721259074 (C), and 6:87183034 (C); for the CSN1S2 gene, rs463985801 (G); and for the CSN3 gene, 6:87390198 (G), 6:87390448 (T), rs110870535 (C), rs716557965 (G), rs43706475 (T), 6:87390604 (T), and rs43703017 (A). The variants rs109299401 (T) and rs43703011 (G) of the CSN1S1 gene, rs441966828 (C) of the CSN1S2 gene, and rs439304887 (A) of the CSN3 gene, which were only observed in the Guabala breed, were excluded.

Table 2 describes the allelic frequencies of polymorphic variants of the casein gene group. In the Guabala population, the SNP rs133474041 of the CSN1S1 gene showed the highest reference allele (G) frequency of 0.800. In the Guaymi population, the RefSNPs rs43703011 (CSN2), rs441966828 (CSN1S2), and rs439304887 (CSN3) all presented the highest frequencies (0.842) of their reference alleles G, C, and A, respectively. The SNP rs43703011 of the CSN2 gene was monomorphic in allele C (1.000) in the Guabala breed and had a frequency of 0.842 in the Guaymi breed. The CSN1S2 gene was polymorphic at rs441966828 in the Guaymi breed, with a C frequency of 0.842. In the case of CSN3, of the 12 alleles evaluated, 5 were polymorphic in the Guaymi breed and 4 in Guabala. The genotypic frequencies by population, considering the polymorphic markers, showed higher values of homozygosity in the Guabala breed over the Guaymi breed, with GG (0.640 and 0.468) at the rs133474041 variant, CC at the rs450402006 variant (0.537 and 0.433), CC at the rs43703015 variant (0.321 and 0.250), AA at the rs43703016 variant, and AA at the rs110014544 variant (0.321 and 0.223), respectively.

Regarding the Fixation Indices or *F* statistics for both populations, the values of  $F_{is}$ ,  $F_{it}$ , and  $F_{st}$  were -0.174, -0.135, and 0.033, respectively, which were not significant.

Table 2. Allelic frequencies of polymorphic variants of casein genes of the Guaymi and Guabala races (reference genome UMD 3.1.1).

Gene CSN1S1	RefSeq rs133474041	Consequence 3'UTR variant	Gua	ibala	Gua	aymi
			Allelic frequency		Allelic frequency	
			0.800 (G)	0.200 (A)	0.684 (G)	0.316 (A)
CSN2	rs109299401	Missense variant	1.000 (T)	0.000 (G)	0.789 (T)	0.211 (G)
CSN2	rs43703011ª	Missense variant	1.000 (C)	0.000 (A)	0.842 (C)	0.158 (A)
CSN1S2	rs441966828	Missense variant	1.000 (C)	0.000 (T)	0.842 (C)	0.158 (T)
CSN3	rs450402006	Missense variant	0.733 (C)	0.267 (T)	0.658 (C)	0.342 (T)
CSN3	rs43703015	Missense variant	0.433 (T)	0.567 (C)	0.500 (T)	0.500 (C)
CSN3	rs43703016	Missense variant	0.429 (C)	0.571 (A)	0.500 (C)	0.500 (A)
CSN3	rs439304887	Synonymous variant	1.000 (A)	0.000 (G)	0.842 (A)	0.158 (G)
CSN3	rs110014544	Synonymous variant	0.433 (G)	0.567 (A)	0.528 (G)	0.472 (A)

<sup>a</sup>Associated with variants A1 (A) and A2 (C) c.245C > A, exon 7 (Gallinat et al. 2013; Kolenda and Sitkowska 2021).

# Discussion

The  $N_{\rm e}$  was lower than those reported by Padilla-Doval et al. (2021) when evaluating SNPs of dairy caseins in the Holstein breed in Colombia. The Shannon Index was slightly higher than that reported by Pashaei et al. (2009) in the Holstein breed (0.110) and lower than that of the native Mazarandanian breed of Pakistan (0.210) were also lower than those reported by Dominguez-Viveros et al. (2021) in Bos taurus breeds such as Hereford and Limousin and by Villalobos-Cortes et al. (2021b) in Guabala and Guaymi populations. The high percentage of monomorphic alleles could be counterproductive since it would reflect the possible presence of homozygous regions by inbreeding, as reflected in the presence of a low number of effective alleles  $(N_e)$ , and where the Guabala breed has the lowest values (1.167). Such behaviour in local breeds with small population sizes is common, and the increase in consanguinity is one of the most relevant problems and entails different negative effects, such as the reduction of phenotypic values (Mastrangelo et al. 2016). This behaviour could be reversed by reorganizing the different mating systems between farms of producers (currently, new breeders organization of Guaymi and Guabala cattle have been identified, with whom new crossbreeding strategies could be developed) and conservation centres or in vitro germplasm banks (FAO 2007). Another alternative that cannot be ruled out would be to implement absorbent crossing strategies with populations with close genetic distances. The allelic frequencies of the SNP rs133474041 are lower than those reported by Kolenda and Sitkowska (2021), who report a frequency of 0.994 for the G variant and 0.061 for the A variant in the Holstein-Friesian breed from Poland. Regarding rs109299401 (CSN2), the T variant of the Guabala breed was more common than that reported by Kolenda and Sitkowska (2021), who described a frequency of 0.930. In the Guaymi population, it was lower, at 0.789. Both values of Guaymi and Guabala SNP rs43703011 of the CSN2 gene are higher than those reported by Kolenda and Sitkowska (2021) and those reported by Bisutti et al. (2022) in Holstein cattle, with a frequency in C of 0.560. This group of alleles has been of growing interest because some studies suggest it may produce intolerance and gastrointestinal problems (Jiangin et al. 2016; Nuhriawangsa et al., 2021) and type 1 diabetes mellitus in infants (Elliott et al. 1999; Chia et al. 2017) and ischaemic heart disease in adults (McLachlan 2001), associated with the release of beta-casomorphin-7 by the presence of histidine (His67) in the A1A1 variant, unlike

the A2A2 variant, which is associated with health benefits, Brooke-Taylor et al. (2017), although some evidence goes against this (Venn et al. 2005; Cass et al. 2008). The difference between the two alleles results in an amino acid difference. The original codon CCT, which codes for the amino acid proline in variant A2, mutates to CAT, forming histidine, in variant A1 at position 67 of CSN2 (Bâlteanu et al. 2010; Oleński et al. 2012). Allele A2 represents the original gene of the genus Bos. This gene encodes the A2 allelic form of betacasein and is present in the milk of many mammals, such as humans, sheep, goats, and bovines (Ng-Kwai-Hang and Grosclaude 2003). The rs441966828 locus in CSN1S2 gene was lower than the values reported by Vanvanhossou et al. (2021) in the African breeds Lagune and Somba (1000) and the crosses of Borgou and Pabli (0.980). Likewise, Meier et al. (2019) reported monomorphic alleles in C of the German Black Pied and Holstein-Friesian populatios. The frequencies of allele C at rs450402006 in Guaymi (0.658) and Guabala (0.733) were lower than the 0.939 reported by Kolenda and Sitkowska (2021). As for the T allele of the CSN3 gene and rs43703015, the frequencies reported in this study are lower than those obtained in Germany by Meier et al. (2019) in populations German Black Pied (0.867) and Holstein-Friesian (0.797) and in the Holstein-Friesian breed (0.992) by Kolenda and Sitkowska (2021) and similar to those reported by Vanvanhossou et al. (2021) in Benin and Nigeria (0.500).

The frequencies of the C allele of the SNP rs43703016 of the creole breeds in this study were lower than those obtained in the Holstein-Friesian populations in Germany (Kolenda and Sitkowska 2021) but similar to those reported in the Lagune and Somba breeds in Benin (Vanvanhossou et al. 2021). The frequency of A (1.000) in the SNP rs439304887 in the Guabala breed was equal to that obtained in the Holstein-Friesian breed of Poland (Kolenda and Sitkowska 2021), while in Guaymi, an allele C frequency equal to 0.842 was observed. At the SNP rs110014544 of CSN3, the breeds had G allele frequencies of 0.433 (Guabala) and 0.528 (Guaymi), which were slightly higher than those seen in the dairy breeds cited above. These high levels of homozygosis in both populations can be attributed to the small number of these breeds that are within conservation programmes. Population censuses as reported by Delgado et al. (2018) in Panama estimate that the Guaymi breed represents 0.08% and the Guabala 0.05% of the livestock population. It is necessary to continue working on models of crosses and conservation modalities whose strategies consider the implementation of conservation centres with in situ, in vivo, and in vitro modalities. It is also vital to promote the creation of breeders' associations that ensure the sustainability of the breeds over time, which would increase the number of animals, prevent inbreeding, and initiate genetic improvement processes, which have not been developed yet. It is also important to consider new analysis tools, such as genomic evaluation, to determine with greater precision the population structures of these landraces their presence of homozygous segments, and genomic inbreeding, among other analyses (Kardos et al. 2015). When we consulted the polymorphic variants in Cattle QTLdb, four of them, two in the CSN2 gene and two in the CSN3 gene, were positive for traits of economic interest. In the CSN2 gene, the rs109299401 variant has been associated with somatic cell count, longevity, milk yield, and protein yield, the rs43703011 variant with somatic cell count, longevity, fat yield, and protein yield. In the CSN3 gene, the rs43703015 variant has been associated with curd firmness and fat vield, the rs43703016 variant with protein percentage (Schopen et al. 2011; Fontanesi et al. 2014; Viale et al. 2017).

# Conclusion

This study determined for the first time, the genetic diversity of the casein group in the Guaymi and Guabala populations, which had few polymorphic alleles, particularly Guabala. Genotypic and allelic frequencies for Guaymi and Guabala cattle were similar to those reported in several Bos taurus breeds. Both breeds had a high prevalence of the A2A2 genotype at the rs43703011 allele of CSN2, which is considered favourable to produce good quality milk in both breeds. With the emergence of a new association of Guaymi and Guabala cattle breeders in Panama (ACCRIPA) and the results obtained in this work. it is proposed to redesign a mating system that includes these new herds. This will allow greater efficiency in conservation programmes, a reduction in inbreeding generated by the low number of animals, in addition to taking advantage of them for commercial purposes, such as the production of type A2A2 milk.

## Acknowledgements

We thank the University of Wagenigen for the support in the receipt and analysis of the DNA samples, in particular to Dr Richard Crooijmans for his help with the receipt of the samples. We thank the Agricultural Innovation Institute of Panama (IDIAP), the National Secretariat of Science, Technology, and Innovation of Panama (SENACYT), and the National Research System (SNI) for their support in this research.

#### **Disclosure statement**

No potential conflict of interest was reported by the author(s).

# Funding

This work was partially funded by the European Union's Horizon 2020 research and innovation programme under grant agreement No. 677353, as well as partially by funds from the National Research System of SENACYT.

# ORCID

Axel Villalobos-Cortés D http://orcid.org/0000-0003-4223-0560

#### References

- Alexander LJ, Stewart AF, MacKinlay AG, Kapelinskaya TV, Tkach TM, Gorodetsky SI. 1988. Isolation and characterization of the bovine kcasein gene. Eur J Biochem. 178:395–401.
- Alim MA, Dong T, Xie Y, Wu XP, Zhang Y, Zhang S, Sun DX. 2014. Effect of polymorphisms in the CSN3 (κ-casein) gene on milk production traits in Chinese Holstein cattle. Mol Biol Rep. 41(11):7585–7593.
- Asmarasari S, Sumantri C, Gunawan A, Taufik E, Anggraeni A. 2020. Genetic variants of milk protein genes and their association with milk components in Holstein Friesian cattle. Jurnal Ilmu Ternak Dan Veteriner. 25(3):99–111.
- Balteanu VA, Vlaic A, Suteu M, Carsai TC. 2010. A comparative study of major milk proteinpolymorphism in six Romanian cattle breeds. Bulletin of University of Agricultural Sciences and Veterinary Medicine Cluj-Napoca—Animal Science and Biotechnologies, 67:345–350. doi:10. 15835/buasvmcn-asb:67:1-2:5322
- Belkhir K, Borsa P, Chikhi L, Raufaste N, Bonhomme F. 2003. Genetix v. 4.04, Logiciel sous WindowsTM pour la Génétique des Populations. Laboratoire Génome et Population, Université Montpellier 2, Montpellier. See http://www.univmontp2.fr/~genetix/genetix/genetix.htm
- Bisutti V, Pegolo S, Giannuzzi D, Mota LFM, Vanzin A, Toscano A, Trevisi E, Ajmone Marsan P, Brasca M, Cecchinato A. 2022. The B-casein (CSN2) A2 allelic variant alters milk protein profile and slightly worsens coagulation properties in Holstein cows. J. Dairy Sci. 105:3794–3809.
- Brooke-Taylor S, Dwyer K, Woodford K, Kost N. 2017. Systematic review of the gastrointestinal effects of A1 compared with A2 β-casein. Adv Nutr. 8(5):739–748. PMID: 28916574; PMCID: PMC5593102.
- Cass H, Gringras P, March J, McKendrick I, O'Hare AE, Owen L, Pollin C. 2008. Absence of urinary opioid peptides in children with autism. Arch Dis Child. 93(9):745–750. doi:10.1136/adc.2006.11. PMID: 18337276.
- Cezard T, Cunningham F, Hunt SE, Koylass B, Kuma N, Saunders G, Shen A, Silva AF, Tsukanov K, Venkataraman S, et al. 2021. The European variation archive: a FAIR resource of genomic variation for all species. Nucleic Acids Res 50:D1216–1220.
- Chia JSJ, McRae JL, Kukuljan S, Woodford K, Elliott RB, Swinburn B, Dwyer KM. 2017. A1 beta-casein milk protein and other environmental pre-disposing factors for type 1 diabetes. Nutr Diabetes. 7:e274.
- Cieslinska A, Kostyra E, Kostyra H, Olenski K, Fiedorowicz E, Kaminski S. 2012. Milk from cows of different β-casein genotypes as a source of βcasomorphin-7. Int J Food Sci Nutr. 63:426–430.
- Delgado JV, Camacho M, Benavente M. 2018. FCPT (FAO/CONBI census projection tool). Una herramienta para estimar los censos raciales nacionales. Acta Iberoam Conserv Anim. 12:179–184.
- Delgado JV, Martinez AM, Acosta A, Alvarez LA, Armstrong E, Camacho E, Canon J, Cortes O, Dunner S, Landi V, et al. 2011. Genetic characterization of Latin-American creole cattle using microsatellite markers. Anim Genet. 43:2–10.
- Dominguez-Viveros J, Medellin-Cazares A, Aguilar-Palma N, Jahuey-Martinez FJ, Rodriguez-Almeida FA. 2021. Definición y analisis del panel de polimorfismos de nucleótido simple a utilizar en pruebas de paternidad para tres razas de bovinos. Rev mex cienc pecu. 12(3):987– 995.
- Eenennaam A, Medrano JF. 1991. Milk protein polymorphisms in California dairy cattle. J Dairy Sci. 74:1730–1742.
- Eigel WN, Butler JE, Ernstrom CA, Farrell HM, Halwarkar VR, Jenness R, Whitney RM. 1984. Nomenclature of proteins of cow's milk: fifth revision. J Dairy Sci. 87:1641–1674.
- Elliott RB, Harris DP, Hil JP, Bibby NJ, Wasmuth HE. 1999. Type I (insulindependent) diabetes mellitus and cow milk: casein variant consumption. Diabetologia. 42(3):292–296.
- Elsik CG, Unni DR, Diesh CM, Tayal A, Emery ML, Nguyen HN, Hagen DE. Bovine Genome Database: new tools for gleaning function from the Bos taurus genome. Nucleic Acids Res. 2016 Jan 4;44(D1):D834-9. doi:10.1093/nar/gkv1077. Epub 2015 Oct 19. PMID: 26481361; PMCID: PMC4702796

- Excoffier L, Laval J, Schneider S. 2005. Arlequín ver. 3.0: An integrated software package for population genetics data analysis. Evol Bioinform Online 1: 47–50.
- FAO. 2007. La Situación de los Recursos Zoogenéticos Mundiales para la Alimentación y la Agricultura –resumen, editado por Dafydd Pilling & Barbara Rischkowsky. Roma.
- Ferretti L, Leone P, Sgaramella V. 1990. Long range restriction analysis of the bovine casein genes. NucleicAcids Res. 18:6829–6833.
- Fontanesi L, Calò DG, Galimberti G, Negrini R, Marino R, Nardone A, Ajmone-Marsan P, Russo V. 2014. Candidate gene association study for nine economically important traits in Italian Holstein cattle. Anim Genet. 45(4):576–580.
- Gallinat JL, Qanbari S, Drögemüller C, Pimentel EG, Thaller G, Tetens J. 2013. DNA-based identification of novel bovine casein gene variants. J Dairy Sci. 96:699–709.
- Ginja C, Gama LT, Cortés O, Martin Burriel IA, Vega-Pla JL, Penedo C, Sponenberg P, Cañón J, Sanz A, et al. 2019. The genetic ancestry of American creole cattle inferred from uniparental and autosomal genetic markers. Sci Rep. 9:11486.
- Guo SW, Thompson EA. 1992. Performing the exact test of Hardy-Weinberg proportion for multiple alleles. Biometrics. 48:361–372.
- Hayes H, Petit E, Bouniol C, Popesc P. 1993. Localization of the α-S2-casein gene (CASAS2) to the homoeologous cattle, sheep, and goat chromosomes 4 by in situ hybridization. Cytogenet Cell Genet. 64:281–285.
- Howe KL, Achuthan P, Allen J, Allen J, Alvarez-Jarreta J, Amode MR, Armean IM, Azov AG, Bennett R, Bhai J, et al. 2021. Ensembl 2021. Nucleic Acids Res. 49(D1):D884–D891.
- Jianqin S, Leiming X, Lu X, Yell G, Ni J, Clarke AJ. 2016. Effects of milk containing only A2 beta casein versus milk containing both A1 and A2 beta casein proteins on gastrointestinal physiology, symptoms of discomfort, and cognitive behavior of people with self-reported intolerance to traditional cows' milk. Nutr J. 15:35.
- Kaminski S, Cieslinska A, Kostyra E. 2007. Polymorphism of bovine betacasein and its potential effect on human health. J Appl Genet. 48:189– 198.
- Kardos M, Luikart G, Allendorf F. 2015. Measuring individual inbreeding in the age of genomics: marker-based measures are better than pedigrees. Heredity. 115:63–72.
- Kolenda M, Sitkowska B. 2021. The polymorphism in various milk protein genes in Polish Holstein-Friesian dairy cattle. Animals (Basel). 11:389.
- Kost NV, Sokolov ÎY, Kurasova ÎB, Dmitriev AD, Tarakanova JN, Gabaeva ÌV, Mikheeva IG. 2009. β-Casomorphins-7 in infants on different type of feeding and different levels of psychomotor development. Peptides. 30:1854–1860.
- Kučerova J, Matějiček A OMJ, Sørensen P, Němcova E, Štipkova M, Kott T, Bouška J, Frelich J. 2006. Milk protein genes CSN1S1, CSN2, CSN3, LGB and their relation to genetic values of milk production parameters in Czech Fleckvieh. Czech J Anim Sci. 51:241–247.
- Laible G, Smolenski G, Wheeler T, Brophy B. 2016. Increased gene dosage for  $\beta$  and  $\kappa$ -casein in transgenic cattle improves milk composition through complex effects. Sci Rep. 6:37607.
- Lischer HE, Excoffier, L. 2012. PGDSpider: an automated data conversion tool for connecting population genetics and genomics programs. Bioinformatics 28(2): 298–299. doi:10.1093/bioinformatics/btr642
- Mastrangelo S, Tolone M, Gerlando RD, Fontanesi L, Sardina MT, Portolano B. 2016. Genomic inbreeding estimation in small populations: evaluation of runs of homozygosity in three local dairy cattle breeds. Animal Consortium 10:746–54. doi:10.1017/S1751731115002943
- McLachlan C. 2001. β-casein A1, ischaemic heart disease mortality, and other illnesses. Med Hypotheses. 56(2):262–272.
- Martien AM, Groenen J, van der Poel J. 1994. Regulation of expression of milk protein genes: a review. Livest Prod Sci. 38(2):61–78.
- Meier S, Korkuć P, Arends D, Brockmann GA. 2019. DNA sequence variants and protein haplotypes of casein genes in German black pied cattle (DSN). Front Genet. 10:1129.

- Ng-Kwai-Hang KF, Grosclaude F. 2003. Genetic polymorphism of milk proteins. In: Fox PF, McSweeney PLH, editors. Advanced dairy chemistry: volume 1: proteins. Boston (MA): Springer; p. 739–816.
- Nilsen H, Olsen HG, Hayes B, et al. 2009. Casein haplotypes and their association with milk production traits in Norwegian red cattle. Genet Sel Evol. 41:24.
- Nuhriawangsa A, Mulyani T, Pambuko G, Vanessa R, Purwadi, Widyas N, Prastowo S. 2021. Molecular screening of bovine β-casein (CSN2) A2 variant in Indonesian Holstein dairy cattle as attempt to produce digestive friendly milk. IOP Conf Ser Earth Environ Sci. 902:012060.
- Oleński K, Cieślińska A, Suchocki T, Szyda J, Kamiński S. 2012. Polymorphism in coding and regulatory sequences of beta-casein gene is associated with milk production traits in Holstein-Friesian cattle. Anim Sci Pap Rep. 30(1):5–12.
- Padilla-Doval J, Zambrano-Arteaga JC, Echeverri-Zuluaga JJ, López-Herrera A. 2021. Analisis genético de cinco polimorfismos de nucleótido simple de caseinas lacteas obtenidos con chips genómicos en ganado Holstein de Antioquia, Colombia. Rev Fac Med Vet Zootec. 68 (2):137–149.
- Pashaei S, Azari MA, Hasani S, Khanahmadi A, Rostamzadeh J. 2009. Genetic diversity in Mazandaranian native cattle: a comparison with Holstein cattle, using ISSR marker. Pak J Biol Sci. 12(9):717–721.
- Peakall R, Smouse PE, 2006. GENALEX 6: genetic analysis in Excel. Population genetic software for teaching and research. Mol Ecol Notes 6:288–295.
- Popescu P, Long S, Riggs P, Womack J, Schmutz S, Fries R, Gallagher DS. 1996. Standardization of cattle karyotype nomenclature: report of the committee for the standardization of the cattle karyotype. Cytogenet Cell Genet. 74:259–261.
- Robinson JT, Thorvaldsdóttir H, Winckler W, Guttman M, Lander ES, Getz G, Mesirov JP. 2011. Integrative genomics viewer. Nat. Biotechnol. 29(1): 24–26. doi:10.1038/nbt.1754
- Roginski H, Fuquay JW, Fox PF. 2003. Encyclopedia of dairy sciences. Vols. 1–4. London: Academic Press.
- Schopen GC, Visker MH, Koks PD, Mullaart E, van Arendonk JA, Bovenhuis H. 2011. Whole-genome association study for milk protein composition in dairy cattle. J Dairy Sci. 94(6):3148–3158.
- Vanvanhossou S, Giambra J, Yin T, Brügemann K, Dossa LH, König S. 2021. First DNA sequencing in Beninese indigenous cattle breeds captures new milk protein variants. Genes (Basel). 12(11):1702.
- Venn BJ, Skeaff CM, Brown R, Mann JI, Green TJ. 2005. A comparison of the effects of A1 and A2 beta-casein protein variants on blood cholesterol concentrations in New Zealand adults. Atherosclerosis. 188(1):175–178. PMID: 16298373.
- Viale E, Tiezzi F, Maretto F, De Marchi M, Penasa M, Cassandro M. 2017. Association of candidate gene polymorphisms with milk technological traits, yield, composition, and somatic cell score in Italian Holstein-Friesian sires. J Dairy Sci. 100(9):7271–7281.
- Villalobos-Cortes A, Carbono M, Rodriguez A, Arosemena E, Jaen M. 2021a. Phenotypic characterization of the Guaymi breed in conservation centers of Panama. Afr J Agric Res. 17(6):907–915.
- Villalobos-Cortes A, Castillo H, Murillo M, Gonzalez R. 2021b. Polymorphisms of markers associated with milk quality in creole and cross-border populations. julio-diciembre, 2021. Ciencia Agropecuaria no. 33:32–43.
- Villalobos-Cortes A, Gonzalez R, Murillo M, Castillo H. 2020. Reduced panel of single-nucleotide polymorphisms for biodiversity studies in bovines. Ciencia Agropecuaria no. 31:19–36.
- Weir BS, Cockerham CC. 1984. Estimating F-statistics for the analysis of population structure. Evolution 38: 1358–1370.
- Wright S. 1965. The interpretation of population structure by F-statistics with special regard to systems of mating. Evolution 19:395–420.
- Zhi-Liang Hu, Eric Ryan Fritz and James M. Reecy 2007. AnimalQTLdb: a livestock QTL database tool set for positional QTL information mining and beyond. Nucleic Acids Research 35 (Database issue):D604–D609.